

ILLUMINA HiSeq-2000 ADDRESS TRANSFORM

National Center for Biotechnology Information (NCBI) – National Library of Medicine

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Problem Statement

The SRA Illumina Genome Analyzer loaders rely on the GA and GA II address field convention (*flowcell:lane:tile:x:y*) to determine the order of spots in the run data file and to detect duplicates. To constrain the size of these fields, the SRA loader allows up to a maximum of 24,576 for X and Y. With HiSeq-2000 the addressing that reflected use of a discrete camera field per tile has given way to a continuous camera field covering much more area, so Y values up to 200,000 are commonly seen. All HiSeq-2000 runs have this problem.

There is no simple way to fix the SRA loaders to adapt to HiSeq-2000 data. Instead the SRA loaders will adopt a more general strategy of address determination and duplicate detection. In the meantime, we propose that submitters transform their HiSeq-2000 data in such a way as to restore the GA addressing parameters without loss of data or information. This approach will be reversible if it later becomes possible to archive the native addresses. Runs receiving this treatment should be specially marked. A fix in the SRA loader code will make this solution redundant. The SRA addressing improvement is expected early 2011.

Treatment

We have defined the following spot address transformation:

$$\text{Spot_Tile*} = (\text{Spot_Tile} \times 100) + (\text{Spot_Y} / 20000)$$

$$\text{Spot_Y*} = \text{Spot_Y} \% 20000$$

Along with this transformation, some of the transformed spots need to be relocated to later in the *qseq* files to ensure that tile addresses are contiguous, which is another requirement of the SRA loaders. For example, in a file that we received from a submitter *s_1_1_0001_qseq.txt*, is the following sequence of spots (line numbers indicated on the left):

242328:	SL-HAG	118	1	1	2204	19980
242329:	SL-HAG	118	1	1	2213	19996
242330:	SL-HAG	118	1	1	2187	20000
242331:	SL-HAG	118	1	1	2295	19756
242332:	SL-HAG	118	1	1	2380	19756

The address transformation will have this result:

242328:	SL-HAG	118	1	100	2204	19980
242329:	SL-HAG	118	1	100	2213	19996
242330:	SL-HAG	118	1	101	2187	0
242331:	SL-HAG	118	1	100	2295	19756
242332:	SL-HAG	118	1	100	2380	19756

In order for the transformed address on record 242330 to load successfully, it is relocated further into the file as record 245435:

242328:	SL-HAG	118	1	100	2204	19980
242329:	SL-HAG	118	1	100	2213	19996
242330:	SL-HAG	118	1	100	2295	19756
242331:	SL-HAG	118	1	100	2380	19756
...						
245435:	SL-HAG	118	1	101	2187	0

To accomplish this reordering, the transform must buffer spots that are assigned to a tile (101) not currently being input (100). Then, as the tile value changes on input (to 101), the currently buffered spots (tile 101) are written out and the cycle is repeated.

A perl script is provided below that performs this operation on *qseq* files that should be available in your Illumina run folder.

Please also add a RUN_ATTRIBUTE, *HISeq_address_transform*, with a value of *yes*, to the Run xml in order to record the application of the address transformation.

The SRA submission can contain either the modified *qseq* file(s) (filetype is fastq), or can be converted into SRF format using the *illumina2srf* utility from *sequenceread-2.1.2* (<http://sourceforge.net/projects/sequenceread>) (be sure to use version 2.1.2 or later, as *sequenceread-2.1.1* had problems with the modified tiles (e.g. a tile of 100 changes to 1)).

Example

A screen capture of the modified and loaded run SRR064189 is presented below.

Run Browser: Browse: Sequence Read Archive: NCBI/NLM/NIH - Mozilla Firefox

File Edit View History Bookmarks Tools Help

http://www.ncbi.nlm.nih.gov/Traces/sra/sra.cgi?cmd=viewer&m=data&s=viewer&run=SRR06

Run Browser: Browse: Sequence Re...

Studies Samples Analyses **Run Browser** Provisional SRA

Run Browser

Experiment: [SRX025768](#)
 1000 Genomes Low Coverage Sequencing - Puerto Rican Population

Run:

Accession: SRR064189
 Alias: BI.PE.100706_SL-HAG_0118_BFC207H4ABXX.1
 Instrument model:
 Date of run: 2010-07-06 04:00:00
 Run center: BI

Other:

Study: [Low coverage of the Puerto Rican in Puerto Rico](#)
 Design: 1000 Genomes Low Coverage Sequencing - Puerto Rican Population
 Platform: ILLUMINA
 Sample: [Human 1000 genomes individual HG00551](#)
 Library Name: Solexa-34369
 Library Strategy: WGS
 Library Source: GENOMIC
 Library Selection: RANDOM
 Library Layout: PAIRED (NOMINAL_SDEV=68.518, NOMINAL_LENGTH=550, ORIENTATION=5'3'-3'5')

Statistics:

Number of spots: 94792104
 Number of reads: 189584208

Find spots: X: Y: View: reads ([customize](#))

[What can the filter be applied to?](#)

< 1 24544 9479211 >

Reads (separated)

245431. [SRR064189.245431](#)
 name:SL-HAG_118:1:100:21397:19953
 x:21397, y:19953

245432. [SRR064189.245432](#)
 name:SL-HAG_118:1:100:21263:19975
 x:21263, y:19975

245433. [SRR064189.245433](#)
 name:SL-HAG_118:1:100:21353:19985
 x:21353, y:19985

245434. [SRR064189.245434](#)
 name:SL-HAG_118:1:100:21305:19988
 x:21305, y:19988

245435. [SRR064189.245435](#)
 name:SL-HAG_118:1:101:2187:0
 x:2187, y:0

```
>gnl|SRA|SRR064189.245435.1 SL-HAG_118:1:101:2187:0 Application Read (Forward)
TCATTTGTAGGGAGCTCACAGTCCTAATATCATGCTTTTGACCTTAGCTTATATCTACA
GTTTAATAATGAATTCCTTGTGACTAGCTGACTTAGGAAAA

>gnl|SRA|SRR064189.245435.2 SL-HAG_118:1:101:2187:0 Application Read (Reverse)
CTGGCCAGGCACAGTGGCTCACACCTGTAATCCCAGCACTTTGGGAAGCTGAGCAGTGTG
AAGCACTACTTGCCAGGAGTTTGAACCCACGCTGGTCAACA
```

Done

To view this interactively, please visit :

<http://www.ncbi.nlm.nih.gov/Traces/sra/sra.cgi?cmd=viewer&m=data&s=viewer&run=SRR064189>

Method

```
#!/opt/perl-5.8.8/bin/perl
# This is a code excerpt that performs address transformation on Illumina HiSeq-2000
reads in order to
# make them loadable by SRA loaders relying on GA and GA II spot addressing
convention.
#

use strict;

die "\nHiSeq2000_2_SRA.pl < qseq file >\n\n"
    if ( scalar @ARGV eq 0 && -t STDIN );

my $TILE_INDEX = 3;
my $Y_INDEX = 5;

# Initialize using first line in qseq file

my $firstLine = <>;
my @spot = split(/\t/, $firstLine);
adjustSpot ( \@spot );
printSpot ( \@spot );
my $currTile = $spot[$TILE_INDEX];
my $nextTile = 0;
my @nextTileSpots = ();

# Continue to process qseq file

while (<>)
{
    my @spot = split(/\t/, $_);
    adjustSpot ( \@spot );

    # Print spots in *$currTile*

    if ( $spot[$TILE_INDEX] eq $currTile )
    {
        printSpot ( \@spot );
    }

    # Determine first *$nextTile* value and start collecting
    # *$nextTile* spots into @nextTileSpots .

    elsif ( ! ( $nextTile ) )
    {
        $nextTile = $spot[$TILE_INDEX];
        push @nextTileSpots, \@spot;
    }

    # After *$nextTile* is set, continue collecting *$nextTile*
    # spots into @nextTileSpots

    elsif ( $spot[$TILE_INDEX] eq $nextTile )
    {
        push @nextTileSpots, \@spot;
    }

    # If *$spot[$TILE_INDEX]* is not *$currTile* or *$nextTile*,
    # then set *$currTile* and *$nextTile* to new values.
    # Output spots collected in @nextTileSpots, and start
```

```

# collecting a new set of *$nextTile* spots in @nextTileSpots.

else
{
    printTileSpots ( \@nextTileSpots );
    $currTile = $nextTile;
    $nextTile = $spot[$TILE_INDEX];
    @nextTileSpots = ();
    push @nextTileSpots, \@spot;
}

}

printTileSpots ( \@nextTileSpots );

#####
sub printTileSpots {
    my $nextTileSpotsRef = shift;
    foreach my $spotRef ( @$nextTileSpotsRef )
    {
        printSpot ( $spotRef );
    }
}

#####
sub adjustSpot {
    my $spotRef = shift;
    $$spotRef[$TILE_INDEX] = ( $$spotRef[$TILE_INDEX] * 100 ) + int (
    $$spotRef[$Y_INDEX] / 20000 );
    $$spotRef[$Y_INDEX] = $$spotRef[$Y_INDEX] % 20000 ;
}

#####
sub printSpot {
    my $spotRef = shift;
    print join ( "\t", @$spotRef );
}

```